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Review

The diagnosis and classification of Henoch–Schönlein purpura: An updated review [☆]Yao-Hsu Yang ^a, Hsin-Hui Yu ^a, Bor-Luen Chiang ^{b,*}^a Department of Pediatrics, National Taiwan University Hospital, College of Medicine, National Taiwan University, Taipei, Taiwan^b Department of Medical Research, National Taiwan University Hospital, Graduate Institute of Clinical Medicine, College of Medicine, National Taiwan University, Taipei, Taiwan

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ABSTRACT

Henoch–Schönlein purpura (HSP) is a common childhood systemic vasculitis with clinical characteristics of cutaneous palpable purpura, arthralgia/arthritis, bowel angina, and hematuria/proteinuria. HSP is identified mainly based on the above presentations. Combined with pathohistological findings of leukocytoclastic vasculitis (LCV) and IgA-immune deposits in vessel walls and/or glomeruli increase the diagnostic sensitivity and specificity. However, considering the accessibility of biopsy and some patients with atypical presentations, there are still medical unmet needs in HSP diagnosis. This article reviews the diagnosis of HSP including the aspects of classification criteria, differential diagnosis, and some laboratory findings as the biomarkers with diagnostic potential.

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1. Introduction

Henoch–Schönlein purpura (HSP) is a systemic small vessel vasculitis that occurs commonly in children. The annual incidence is 13–20 per 100,000 children under 17 years old [1,2]. It is characterized by non-thrombocytopenic palpable purpura that mostly located on the dependent parts like lower extremities and buttocks, arthralgia/arthritis,

bowel angina, and hematuria/proteinuria. Treatment is supportive because the disease course is usually benign and self-limited [3]. Progressive impairment of renal function, bowel perforation, and central nerve system involvement is rare but major morbidity of HSP [4–6]. Aggressive therapies with steroid and/or immunosuppressants are then indicated under such conditions.

Although this disease is not uncommon in children, the etiology and pathogenesis are still yet to be determined. Previous epidemiological studies have found striking seasonal variations in HSP, with most cases occurring in the autumn and winter. HSP has also been associated with a history of preceding infections, especially upper respiratory tract infection [1,3,7]. In addition, other characteristics of HSP include the deposition of IgA and C3 in small vessel walls, polymorphonuclear neutrophil infiltration around the vessel and in vessel walls, and

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increased serum levels of IgA and proinflammatory cytokines at the acute stage [3,8]. Combined, HSP is regarded as a specific immune-mediated entity induced by environmental factors, particularly infections.

Clinically, since there are no disease-specific laboratory abnormalities, HSP is currently diagnosed based on symptoms and signs and histopathological findings [3,9]. In this article, we reviewed and compared various diagnostic criteria of HSP. In addition, differential diagnosis of HSP and some potential biomarker that may assist in diagnosing HSP were also reviewed.

2. Diagnostic criteria of HSP

Schönlein first described HSP as triad of purpuric rash, arthritis, and abnormalities of the urinary sediment in 1837 [10]. In 1874, Henoch described the association of purpuric rash, abdominal pain with bloody diarrhea, and proteinuria [11]. Up to date, several sets of diagnostic criteria for HSP have been proposed; the detail and comparison of these different classification criteria were further reviewed and discussed as follows.

2.1. The American College of Rheumatology (ACR) criteria

In 1990, ACR first proposed criteria for identifying HSP by comparing 85 patients who had HSP with other 722 patients with other vasculitis. Four criteria were finally identified including palpable purpura not related to thrombocytopenia, age ≤ 20 years at disease onset, acute abdominal pain, and granulocytes in the walls of small arterioles and venules on biopsy (Table 1). A patient shall be diagnosed with HSP if at least 2 of these criteria are present. The presence of any two or more criteria yielded a sensitivity of 87.1% and specificity of 87.7% [9].

2.2. Michel's criteria

By ACR criteria, however, a patient with other vasculitis presents with non-thrombocytopenic palpable purpura and granulocytes in small vessel walls or around vessels on biopsy could be classified as having HSP without other findings. For example, hypersensitivity vasculitis (HV), a kind of leukocytoclastic vasculitis (HCV) that commonly affects

adults [3]. To distinguish between HV and HSP, Michel and co-workers conducted a study comparing 93 patients with HV and 85 patients with HSP and identified 6 criteria: palpable purpura not related to thrombocytopenia, bowel angina, gastrointestinal bleeding, hematuria, age ≤ 20 years at disease onset, and no history of medication intake at disease onset (Table 1). They found 3 or more criteria from the above list of 6 yielded 87.1% of correctly classified HSP cases; and 2 or fewer criteria from the same list of 6 correctly classified 74.2% of HV cases [12].

2.3. Chapel Hill Consensus Conference (CHCC)

Although ACR has proposed classification criteria for HSP that would provide a standard way to evaluate patients with similar presentations, there is no strict uniform definition of this disease. In an attempt to address this problem, a consensus conference on nomenclature of systemic vasculitis was held in Chapel Hill in 1994 [13]. They finally provided a consensus for HSP definition: it is a small vessel vasculitis with IgA-dominant immune deposits, typically involves skin, gut, and glomeruli, and is associated with arthralgia/arthritis (Table 1). This definition was according to the opinion of an expert panel, but was not validated with patient data and was not intended to function as a set of classification criteria [14]. Moreover, IgA-immune vascular deposits are not specific for HSP because they are found in other vasculitis syndromes such as erythema nodosum, cryoglobulinemia, coagulopathic vasculopathies and livedoid vasculitis [15,16].

2.4. Helander's criteria

Many vasculitis like urticarial vasculitis, microscopic polyarteritis nodosa, and collagen vascular disease could be confused as HSP if one uses solely ACR criteria that do not include IgA-immune deposits within small vessels [14]. In 1995, Helander, DeCastro, and Gibson proposed their revised criteria for HSP including cutaneous IgA vascular deposits, age 20 years or younger, gastrointestinal involvement, upper respiratory tract infection prodrome, and renal biopsy showing mesangioproliferative glomerulonephritis with or without IgA deposition (Table 1). The presence of 3 or more of above 5 criteria in patients with palpable purpura and histopathological LCV yielded

Table 1
Summary of classification criteria for Henoch–Schönlein purpura (HSP) diagnosis.

Classification	Diagnostic criteria
ACR 1990 [9]	≥ 2 of the following: 1. Palpable purpura, not thrombocytopenic 2. Bowel angina 3. Wall granulocytes on biopsy 4. Age ≤ 20 years at disease onset
Michel et al. 1992 [12]	≥ 3 of the following: HSP; ≤ 2 of the following: HV 1. Palpable purpura, not thrombocytopenic 2. Bowel angina 3. Gastrointestinal bleeding 4. Hematuria 5. Age ≤ 20 years at disease onset 6. No history of medication intake at disease onset
CHCC 1994 [13]	Vasculitis, with IgA-dominant immune deposits, affecting small vessels (ie, capillaries, venules, or arterioles); typically involves skin, gut, and glomeruli and is associated with arthralgias or arthritis
Helander et al. 1995 [17]	Palpable purpura, not thrombocytopenic with LCV + ≥ 3 of the following: 1. Vascular IgA deposition 2. Age ≤ 20 years at disease onset 3. Gastrointestinal involvement 4. Upper respiratory tract infection prodrome 5. Mesangioproliferative glomerulonephritis with or without IgA deposition
EULAR/PRINTO/PRES 2010 [20]	Palpable purpura, not thrombocytopenic/petechiae (mandatory) + ≥ 1 of the following 1. Diffuse abdominal pain 2. Histopathology: typical LCV with predominant IgA deposits or proliferative glomerulonephritis with predominant IgA deposits 3. Arthritis or arthralgias 4. Renal involvement (proteinuria: >0.3 g/24 h or >30 mmol/mg of urine albumin to creatinine ratio on a spot morning sample; and/or hematuria, red blood cell casts: >5 red cells per high power field or $\geq 2+$ on dipstick or red blood cell casts in the urinary sediment)

ACR, The American College of Rheumatology; HV, hypersensitivity vasculitis; CHCC, Chapel Hill Consensus Criteria; LCV, leukocytoclastic vasculitis; EULAR/PRINTO/PRES, European League Against Rheumatism/Paediatric Rheumatology International Trials Organization/Paediatric Rheumatology European Society.

sensitivity and specificity greater than 90% [17]. However, the skin and kidney biopsies and direct immunofluorescence staining for IgA limit their usage in clinical practice, especially for pediatric patients.

2.5. European League Against Rheumatism/Paediatric Rheumatology International Trials Organization/Paediatric Rheumatology European Society (EULAR/PRINTO/PRES) criteria

Since the above criteria have some limitations [9,12–14,17,18], in 2005 the vasculitis working group of the PRES proposed new classification criteria for pediatric vasculitis including HSP, endorsed by the EULAR. However, these proposed modifications were mainly based on a literature review and a consensus-based process and were not validated. Therefore, EULAR, PRINTO and PRES conducted a statistical validation process in 2008, with a large-scale, web-based data collection [19]. In 2010, the EULAR/PRINTO/PRES criteria for HSP were formally published. The criteria include palpable purpura as a mandatory criterion, together with at least one of the following findings: diffuse abdominal pain, LCV with predominant IgA deposits on skin biopsy, acute arthritis or arthralgias in any joint, and renal involvement as evidenced by proteinuria and/or hematuria (Table 1). The sensitivity and specificity of these classification criteria were 100% and 87% respectively [20].

Comparing with ACR criteria, EULAR/PRINTO/PRES criteria chose histopathology showing typically LCV with predominant IgA deposits or proliferative glomerulonephritis with predominant IgA deposits for all doubtful cases with atypical purpura distribution. Other differences were related to the inclusion of joint and renal involvement [20]. When either set of criteria as applied to their pediatric population, EULAR/PRINTO/PRES criteria showed a better sensitivity and specificity than ACR criteria [14,20]. However, EULAR/PRINTO/PRES criteria were originally derived from data of 823 children with HSP and 356 children with other vasculitis, such criteria have not been validated in adults [19,20].

3. Differential diagnosis of HSP

HSP must be distinguished from other diseases with similar presentations (Table 2). Thrombocytopenic purpura like immune thrombocytopenic purpura can be easily identified by the detection of low platelet

Table 2
Differential diagnosis of Henoch–Schönlein Purpura (HSP).

	Diseases
Thrombocytopenic purpura	Immune thrombocytopenic purpura Thrombotic thrombocytopenic purpura
Other types of vasculitis	Hypersensitivity vasculitis Urticarial vasculitis Mixed cryoglobulinemia Cutaneous polyarteritis ANCA-associated small vessel vasculitis
Rheumatic diseases ^a	Systemic lupus erythematosus Rheumatoid arthritis Sjögren syndrome Mixed connective tissue disorder Juvenile dermatomyositis Antiphospholipid antibody syndrome
Others	Septicemia Disseminated intravascular coagulation Papular-purpuric gloves-and-socks syndrome Mediterranean fever Causes of acute surgical abdomen

ANCA, anti-neutrophil cytoplasmic antibodies.

^a Rheumatic diseases share some symptoms with HSP such as cutaneous purpuric lesions and arthritis.

count. If it is difficult to decide the cause of cutaneous purpuric lesions that may be presented in other types of vasculitis such as urticarial vasculitis, cryoglobulinemia, and HV, a skin biopsy and immunofluorescence study may assist in the diagnosis [3,16,17,20]. Some rheumatic diseases might have presentation of cutaneous vasculitis [3,16,21–25]. Clinical characteristics combined with laboratory abnormalities usually provide assistance to determine the underlying diseases. Furthermore, since abdominal pain is commonly presented in HSP, the more common causes of acute surgical abdomen must be considered [3].

4. Biomarkers for HSP diagnosis

In HSP, the platelet count is normal or increased. A moderate leukocytosis with a left shift is noted in some patients. Antinuclear antibody is mostly undetectable, and serum levels of C3 and C4 are usually within normal limit [3]. Although some proinflammatory cytokines and chemokines are elevated at acute stage [8,26], these laboratory abnormalities are not specific for HSP but can be seen in a variety of inflammatory conditions. The presence of LCV and IgA-immune deposits on skin and/or kidney biopsies significantly increases the diagnostic accuracy for HSP [14,17,20]. However, many vasculitis and glomerulonephritis show similar histopathological findings [14–16,27].

The immune deposits in HSP are principally composed of IgA1 that predominates in serum. IgA1 is structurally different from IgA2 in the hinge region of the heavy chain, where it is rich of proline and composed of 5–6 O-linked glycosylation sites. An abnormal glycosylation of the IgA1 hinge region would occur in the context of a deficiency of galactose and/or sialic acid; such a molecule is prone to cause IgA aggregation and thus macromolecular complexes [8]. Like IgA1 in IgA nephropathy, recent studies have shown aberrant glycosylation of IgA1 in HSP patients with nephritis [28,29]. In addition to the structure of IgA1 in HSP, the search for antigenic epitopes that IgA1 binds to is another important and interesting issue to be addressed. A variety of IgA autoantibodies have been found associated with HSP including IgA rheumatoid factor [30], IgA anticardiolipin antibodies [31,32], and IgA antiendothelial cell antibodies [33,34]. Recently, we found IgA1 of HSP patients bound well to β 2 glycoprotein I (β 2GPI) and some β 2GPI-derived linear peptides. These IgA anti- β 2GPI antibodies were cross-reactive to endothelial cells and induced complement-dependent cell lysis [35]. Although aberrant glycosylated IgA1 and some of above IgA autoantibodies are likely to play a pathogenic role in HSP, whether they are diagnostic biomarkers for HSP needs more studies to validate.

5. Conclusion

The current diagnostic criteria have high sensitivity and specificity; most HSP patients are accurately diagnosed based on them. However, some patients have atypical presentations and biopsies are sometimes not easily obtained. Therefore, the development of less invasive laboratory tests that are of diagnostic value is needed. Clearly, the elucidation of HSP pathogenesis becomes important that may identify some disease-specific biomarkers assisting in HSP diagnosis.

References

- [1] Yang YH, Hung CF, Hsu CR, Wang LC, Chuang YH, Lin YT, et al. A nationwide survey on epidemiological characteristics of childhood Henoch–Schönlein purpura in Taiwan. *Rheumatology (Oxford)* 2005;44:618–22.
- [2] Gardner-Medwin JM, Dolezalova P, Cummins C, Southwood TR. Incidence of Henoch–Schönlein purpura, Kawasaki disease, and rare vasculitides in children of different ethnic origins. *Lancet* 2002;360:1197–202.
- [3] Brogan P, Bagga A. Leukocytoclastic vasculitis. In: Cassidy JT, Petty RE, editors. *Textbook of pediatric rheumatology*. Philadelphia: Saunders, Elsevier Inc.; 2011. p. 483–97.
- [4] Chang WL, Yang YH, Wang LC, Lin YT, Chiang BL. Renal manifestations in Henoch–Schönlein purpura: a 10-year clinical study. *Pediatr Nephrol* 2005;20:1269–72.
- [5] Bissonnette R, Dansereau A, D'Amico P, Patenaude JV, Paradis J. Perforation of large and small bowel in Henoch–Schönlein purpura. *Int J Dermatol* 1997;36:361–3.

- [6] Belman AL, Leicher CR, Moshe SL, Mezey AP. Neurologic manifestations of Schönlein–Henoch purpura: report of three cases and review of the literature. *Pediatrics* 1985;75:687–92.
- [7] Saulsbury FT. Henoch–Schönlein purpura in children. Report of 100 patients and review of the literature. *Medicine (Baltimore)* 1999;78:395–409.
- [8] Yang YH, Chuang YH, Wang LC, Huang HY, Gershwin ME, Chiang BL. The immunobiology of Henoch–Schönlein purpura. *Autoimmun Rev* 2008;7:179–84.
- [9] Mills JA, Michel BA, Bloch DA, Calabrese LH, Hunder GG, Arend WP, et al. The American College of Rheumatology 1990 criteria for the classification of vasculitis. *Arthritis Rheum* 1990;33:1114–21.
- [10] Schönlein JL. *Allgemeine und specielle Pathologie und Therapie*, vol. 2. Herisau, Germany: Literatur-Comptoir; 1837.
- [11] Henoch EH. Über ein eigenthümliche Form von Purpura. *Berl Klin Wochenschr* 1874;11:641–3.
- [12] Michel BA, Hunder GG, Bloch DA, Calabrese LH. Hypersensitivity vasculitis and Henoch–Schönlein purpura: a comparison between the 2 disorders. *J Rheumatol* 1992;19:721–8.
- [13] Jennette JC, Falk RJ, Andrassy K, Bacon PA, Churg J, Gross WL, et al. Nomenclature of systemic vasculitides. Proposal of an international consensus conference. *Arthritis Rheum* 1994;37:187–92.
- [14] Linskey KR, Kroshinsky D, Mihm Jr MC, Hoang MP. Immunoglobulin-A-associated small-vessel vasculitis: a 10-year experience at the Massachusetts General Hospital. *J Am Acad Dermatol* 2012;66:813–22.
- [15] Magro CM, Crowson AN. A clinical and histologic study of 37 cases of immunoglobulin A-associated vasculitis. *Am J Dermatopathol* 1999;21:234–40.
- [16] Carlson JA, Chen KR. Cutaneous vasculitis update: small vessel neutrophilic vasculitis syndromes. *Am J Dermatopathol* 2006;28:486–506.
- [17] Helander SD, De Castro FR, Gibson LE. Henoch–Schönlein purpura: clinicopathologic correlation of cutaneous vascular IgA deposits and the relationship to leukocytoclastic vasculitis. *Acta Derm Venereol* 1995;75:125–9.
- [18] Rao JK, Allen NB, Pincus T. Limitations of the 1990 American College of Rheumatology classification criteria in the diagnosis of vasculitis. *Ann Intern Med* 1998;129:345–52.
- [19] Ruperto N, Ozen S, Pistorio A, Dolezalova P, Brogan P, Cabral DA, et al. EULAR/PRINTO/PRES criteria for Henoch–Schönlein purpura, childhood polyarteritis nodosa, childhood Wegener granulomatosis and childhood Takayasu arteritis: Ankara 2008. Part I: overall methodology and clinical characterization. *Ann Rheum Dis* 2010;69:790–7.
- [20] Ozen S, Pistorio A, Iusan SM, Bakaloglu A, Herlin T, Brik R, et al. EULAR/PRINTO/PRES criteria for Henoch–Schönlein purpura, childhood polyarteritis nodosa, childhood Wegener granulomatosis and childhood Takayasu arteritis: Ankara 2008. Part II: final classification criteria. *Ann Rheum Dis* 2010;69:798–806.
- [21] Marzano AV, Vezzoli P, Berti E. Skin involvement in cutaneous and systemic vasculitis. *Autoimmun Rev* 2013;12:467–76.
- [22] Cardinali C, Caproni M, Bernacchi E, Amato L, Fabbri P. The spectrum of cutaneous manifestations in lupus erythematosus—the Italian experience. *Lupus* 2000;9:417–23.
- [23] Chen KR, Toyohara A, Suzuki A, Miyakawa S. Clinical and histopathological spectrum of cutaneous vasculitis in rheumatoid arthritis. *Br J Dermatol* 2002;147:905–13.
- [24] Ramos-Casals M, Anaya JM, García-Carrasco M, Rosas J, Bové A, Claver G, et al. Cutaneous vasculitis in primary Sjögren syndrome: classification and clinical significance of 52 patients. *Medicine (Baltimore)* 2004;83:96–106.
- [25] Weinstein S, Piette W. Cutaneous manifestations of antiphospholipid antibody syndrome. *Hematol Oncol Clin North Am* 2008;22:67–77.
- [26] Besbas N, Saatci U, Ruacan S, Ozen S, Sungur A, Bakaloglu A, et al. The role of cytokines in Henoch Schönlein purpura. *Scand J Rheumatol* 1997;26:456–60.
- [27] Lai KN. Pathogenesis of IgA nephropathy. *Nat Rev Nephrol* 2012;8:275–83.
- [28] Kiryluk K, Moldoveanu Z, Sanders JT, Eison TM, Suzuki H, Julian BA, et al. Aberrant glycosylation of IgA1 is inherited in both pediatric IgA nephropathy and Henoch–Schönlein purpura nephritis. *Kidney Int* 2011;80:79–87.
- [29] Yu HH, Chiang BL, Yang YH. Altered glycosylation of circulatory IgA1 involved in Henoch–Schönlein purpura and IgA nephropathy. *J Formos Med Assoc* 2012;111:121–2.
- [30] Saulsbury FT. Heavy and light chain composition of serum IgA and IgA rheumatoid factor in Henoch–Schönlein purpura. *Arthritis Rheum* 1992;35:1377–80.
- [31] Yang YH, Huang MT, Lin SC, Lin YT, Tsai MJ, Chiang BL. Increased transforming growth factor-beta (TGF-beta)-secreting T cells and IgA anti-cardiolipin antibody levels during acute stage of childhood Henoch–Schönlein purpura. *Clin Exp Immunol* 2000;122:285–90.
- [32] Kawakami T, Watabe H, Mizoguchi M, Soma Y. Elevated serum IgA anticardiolipin antibody levels in adult Henoch–Schönlein purpura. *Br J Dermatol* 2006;155:983–7.
- [33] Fujieda M, Oishi N, Naruse K, Hashizume M, Nishiya K, Kurashige T, et al. Soluble thrombomodulin and antibodies to bovine glomerular endothelial cells in patients with Henoch–Schönlein purpura. *Arch Dis Child* 1998;78:240–4.
- [34] Yang YH, Wang SJ, Chuang YH, Lin YT, Chiang BL. The level of IgA antibodies to human umbilical vein endothelial cells can be enhanced by TNF-alpha treatment in children with Henoch–Schönlein purpura. *Clin Exp Immunol* 2002;130:352–7.
- [35] Yang YH, Chang CJ, Chuang YH, Hsu HY, Yu HH, Lee JH, et al. Identification and characterization of IgA antibodies against beta-2-glycoprotein I in childhood Henoch–Schönlein purpura. *Br J Dermatol* 2012;167:874–81.